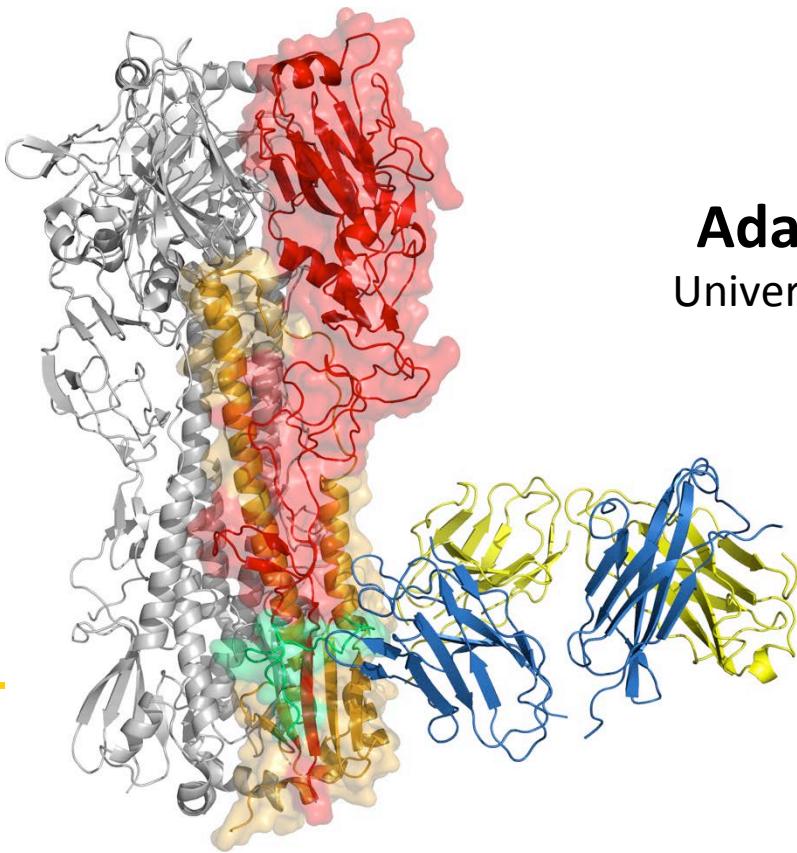
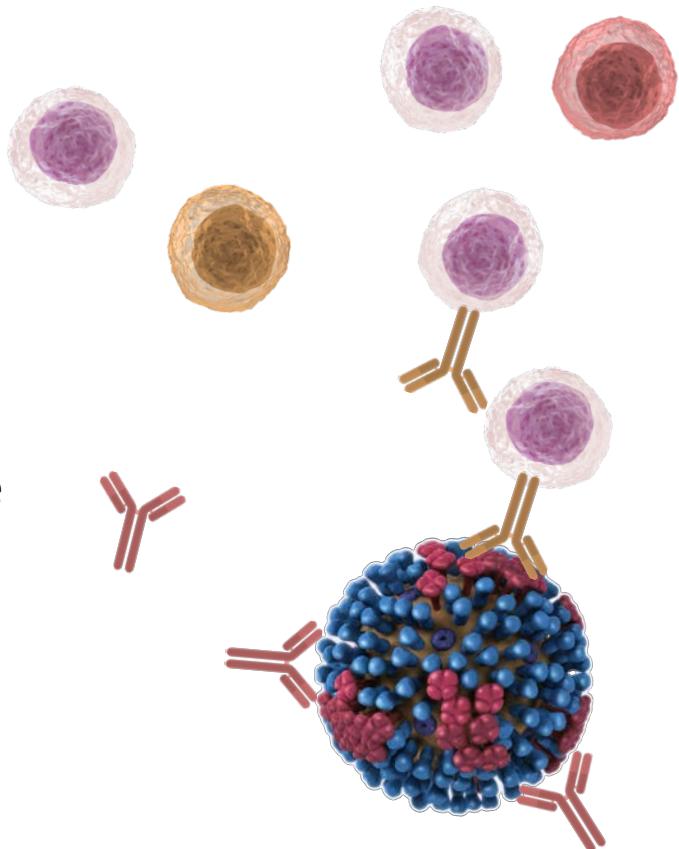


Universal vaccines and monoclonal antibodies



Adam Wheatley
University of Melbourne



Vaccine protection against influenza

Vaccines for epidemic influenza (seasonal)

- Trivalent or quadrivalent formulations
- Inactivated viral preparations mostly HA (IIV3/4)
- Live attenuated? (LAIV3/4)
- **Limitations:**
 - Protection limited to closely matched strains – narrow
 - Variable year to year effectiveness
 - Necessitates influenza surveillance and frequent vaccine reformulation

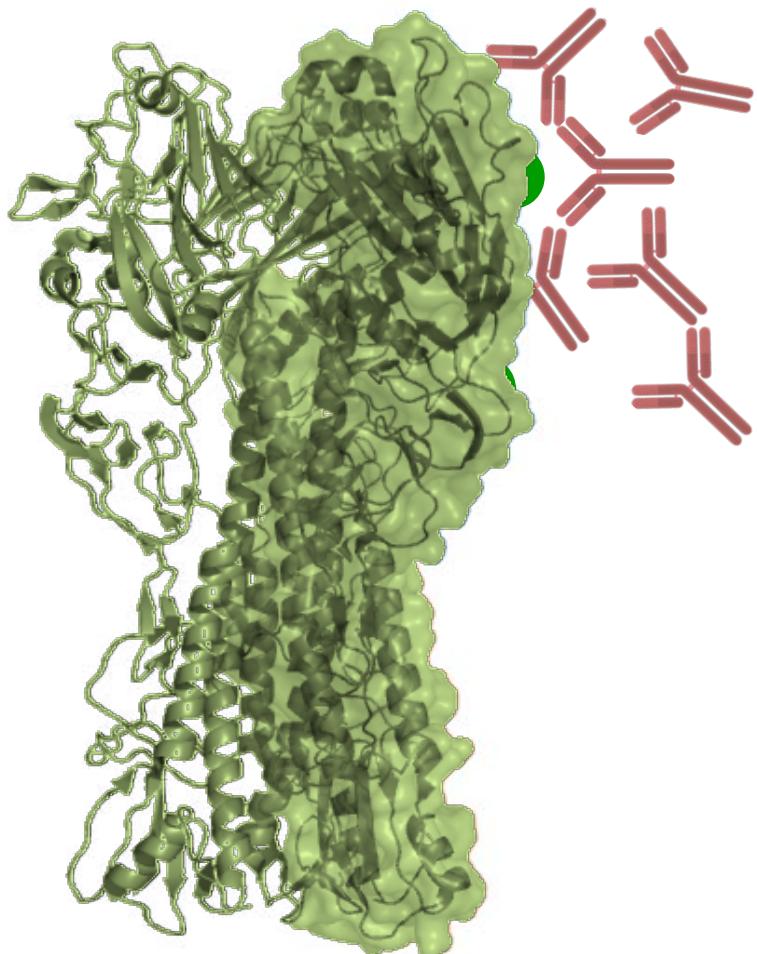
Vaccines for pandemic influenza

- Monovalent formulations against H5N1 or H7N9 strains
- **Limitations:**
 - Long lead times for production → stockpiling
 - Unclear if emergent viruses will match vaccine stocks

So why do current influenza vaccines provide such narrow protection?

The viral hemagglutinin (HA) – a shifting target for antibody

- Immunity preventing acquisition of influenza is primarily mediated by antibody to the major surface proteins - hemagglutinin (HA) and neuraminidase (NA)



- Protective antibodies elicited by infection or vaccination target variable sites surrounding the RBD → blocks viral entry
- Antigenic changes driven by immune pressure leads to a progressive loss of immune recognition (**antigenic drift**)
 - Loss of vaccine effectiveness
- Genetic recombination with environmental viruses (IAV) (**antigenic shift**) and complete loss of immune recognition
 - Loss of vaccine effectiveness

Gamblin et al 2004; PDB ID:1Ruz

Extending influenza vaccine coverage

Influenza B

Yamagata-like

Victoria-like

Pan-IBV

Influenza A

Group 2

H4
H14

H3

H2

H5

Any vaccine that can broaden protection would be a major medical advance

H7

H11

H13
H16

H9

H8

Pan-IAV

Pan-group

H1N1

Pan-subtype

Group 1

0.02

Current
Seasonal
Vaccines

UNIVERSAL VACCINE

Diverse strategies to elicit universal influenza immunity

Common Goal

- Immune responses (vaccines) that better account for antigenic diversity
- Simultaneous protection from both circulating seasonal (antigenic drift) and emerging/pandemic (antigenic shift) influenza strains

1. Cross-reactive T cells

- Antiviral CD4 or CD8 responses targeted to highly conserved viral epitopes
- Limit pathogenesis, viral replication and transmission

2. Antibody responses to conserved viral proteins

- Deriving immunogens from viral proteins with comparatively higher sequence conservation
 - NP, M2, NA

3. Cross-reactive antibody responses to HA

- Identification of highly conserved protective/neutralizing epitopes in HA
- Focusing the humoral responses towards protective neutralising epitopes

A strategy for HA-directed universal vaccines

1. Identify cross-reactive HA epitopes recognised by human antibody/B cells
 - Map epitopes
 - Define the prevalence and ontogeny
 - Define protective breadth, potency and mechanism

2. Reprogram vaccine immunity to target these epitopes
 - Maximise humoral responses targeting cross-reactive epitopes
 - Minimise humoral responses targeting canonical strain-specific epitopes

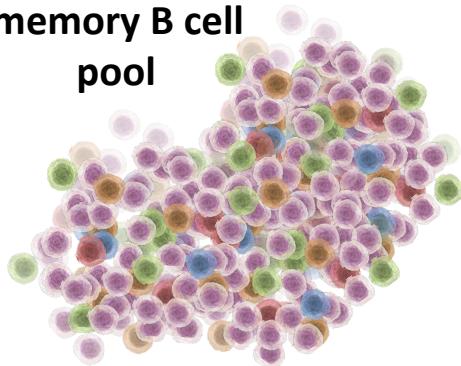
Interrogation of memory B cell pools using recombinant HA probes

Serum antibody

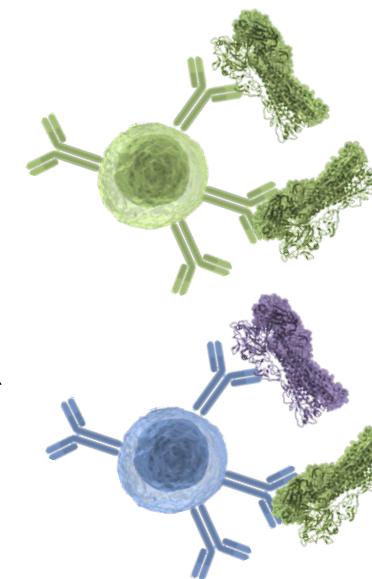
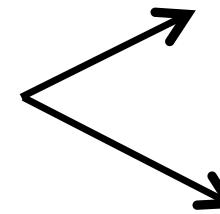
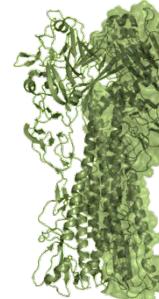


- HI assay – indirect neutralisation at the RBD
- Direct virus neutralisation assays
- Antibody effector functions (ADCC, ADCP)

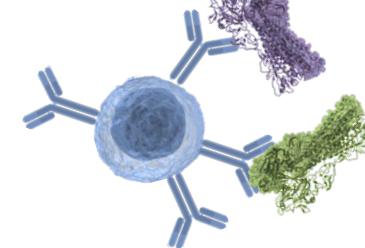
Long lived
memory B cell
pool



Fluorescent
rHA tetramers

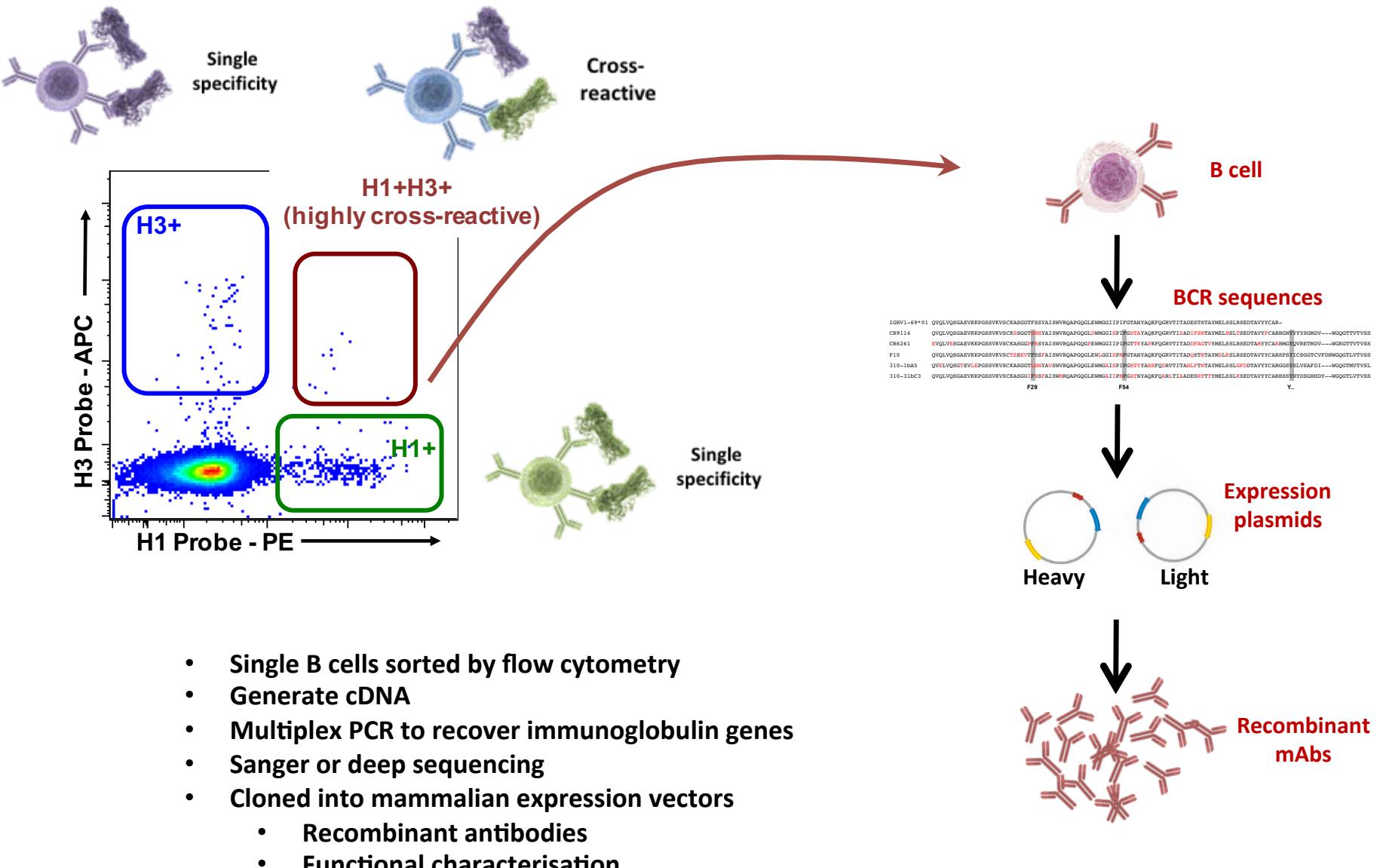


Single
specificity



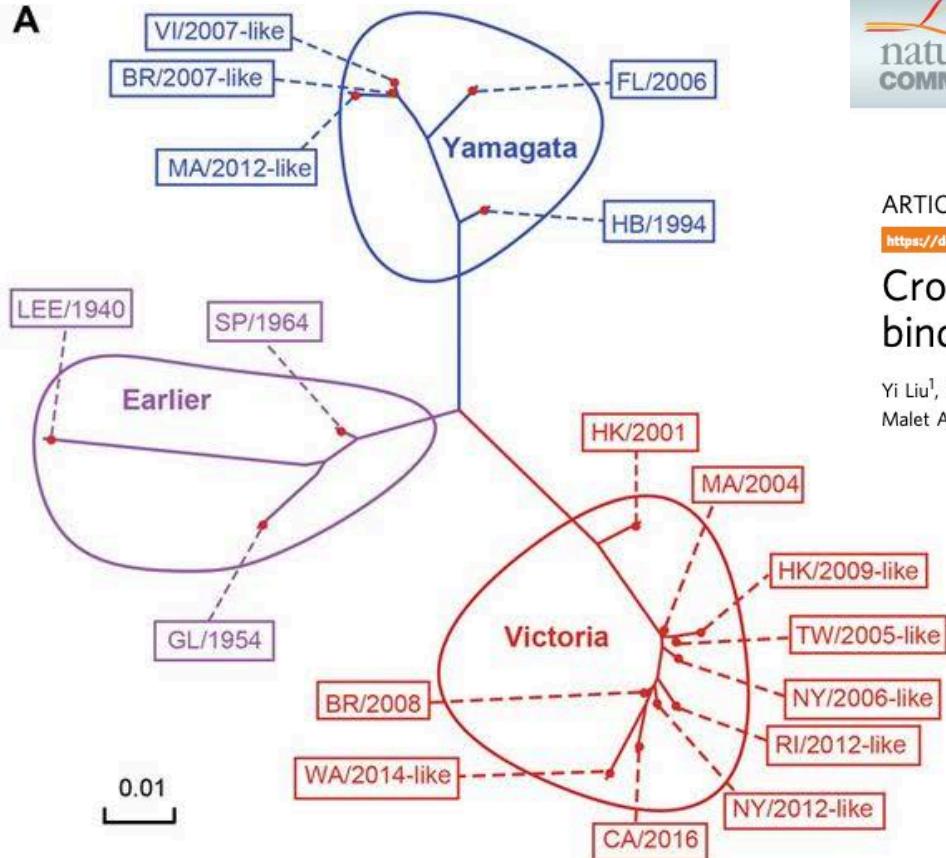
Cross-
reactive

Characterising frequency, phenotype and specificity of HA probe+ B cells



Cross-reactive recognition of influenza B by human antibody

A



ARTICLE

<https://doi.org/10.1038/s41467-018-08165-y>

OPEN

Cross-lineage protection by human antibodies binding the influenza B hemagglutinin

Yi Liu¹, Hyon-Xhi Tan¹, Marios Koutsakos¹, Sinthujan Jegaskanda¹, Robyn Esterbauer¹, Danielle Tilmanis², Malet Aban², Katherine Kedzierska¹, Aeron C. Hurt^{1,2}, Stephen J. Kent^{1,3,4} & Adam K. Wheatley¹

Questions?

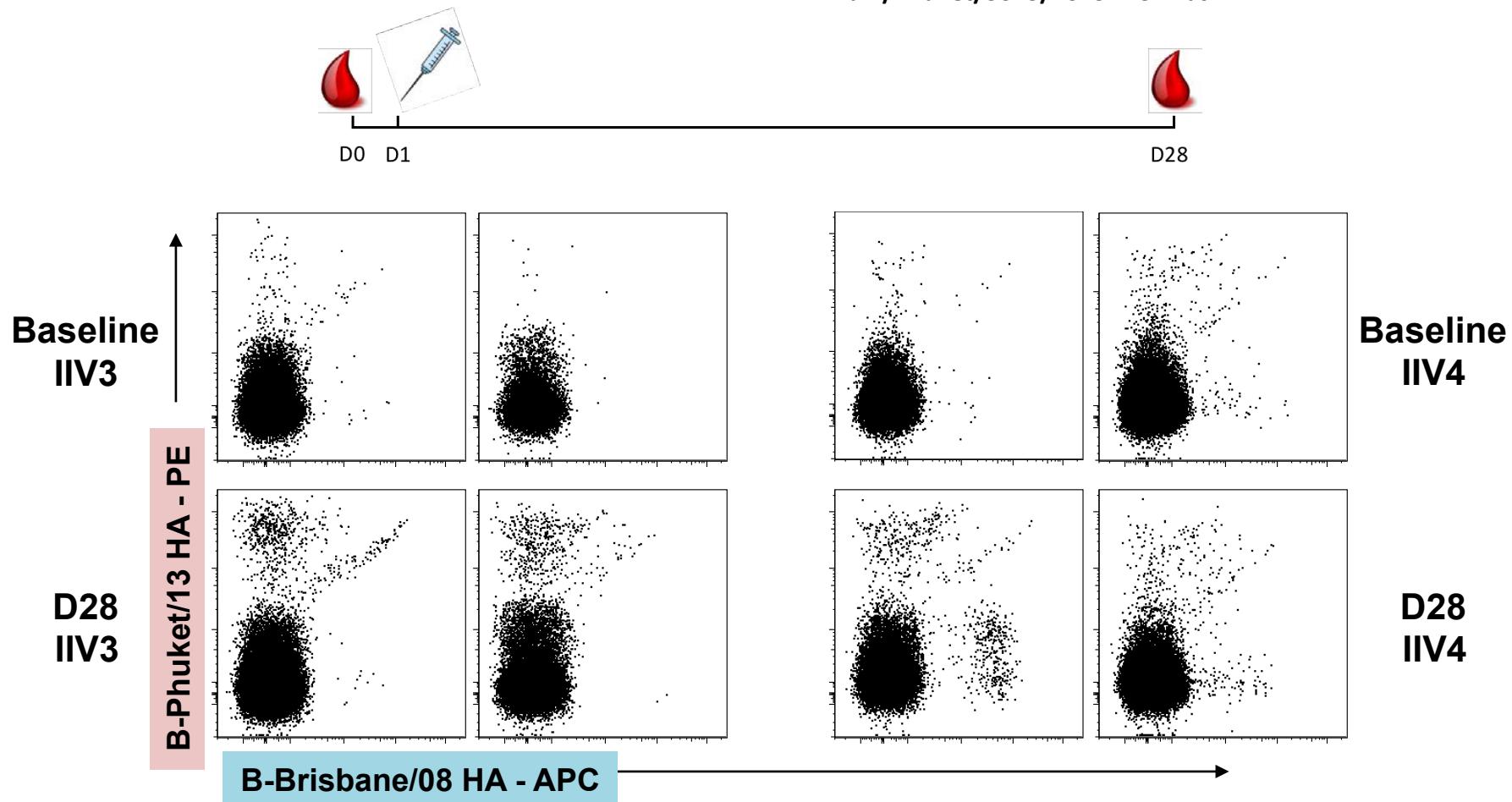
- How much cross-reactivity between the two lineages?
- Is the IBV stem a target?
- IIV3 vs IIV4?

Resolving HA-specific B cell populations in IIV3 and IIV4 cohorts

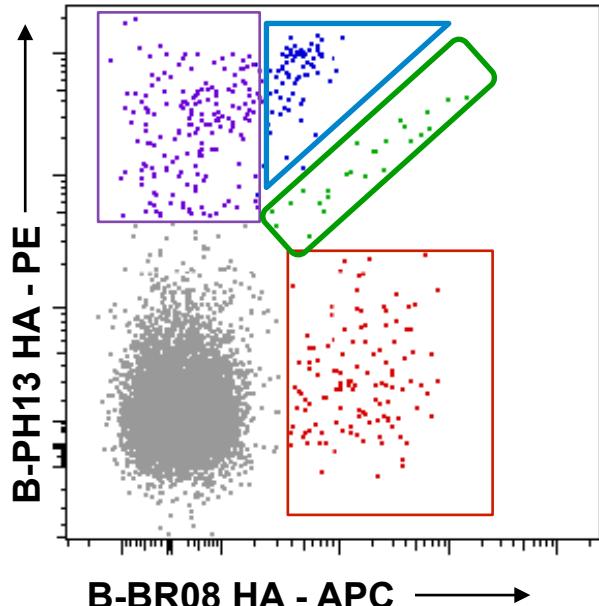
- Recruited 29 subjects (25-56 years old)
- Immunised with 2015 Afluria® (BioCSL)
- Recruited 20 subjects (25-56 years old)
- Immunised with 2016 Fluquadri® (Sanofi)

A (H1N1): an A/California/4/2009 (H1N1)pdm09 like virus
A (H3N2): an A/Switzerland/9715293/2013 (H3N2) like virus
B: a **B/Phuket/3073/2013** like virus.

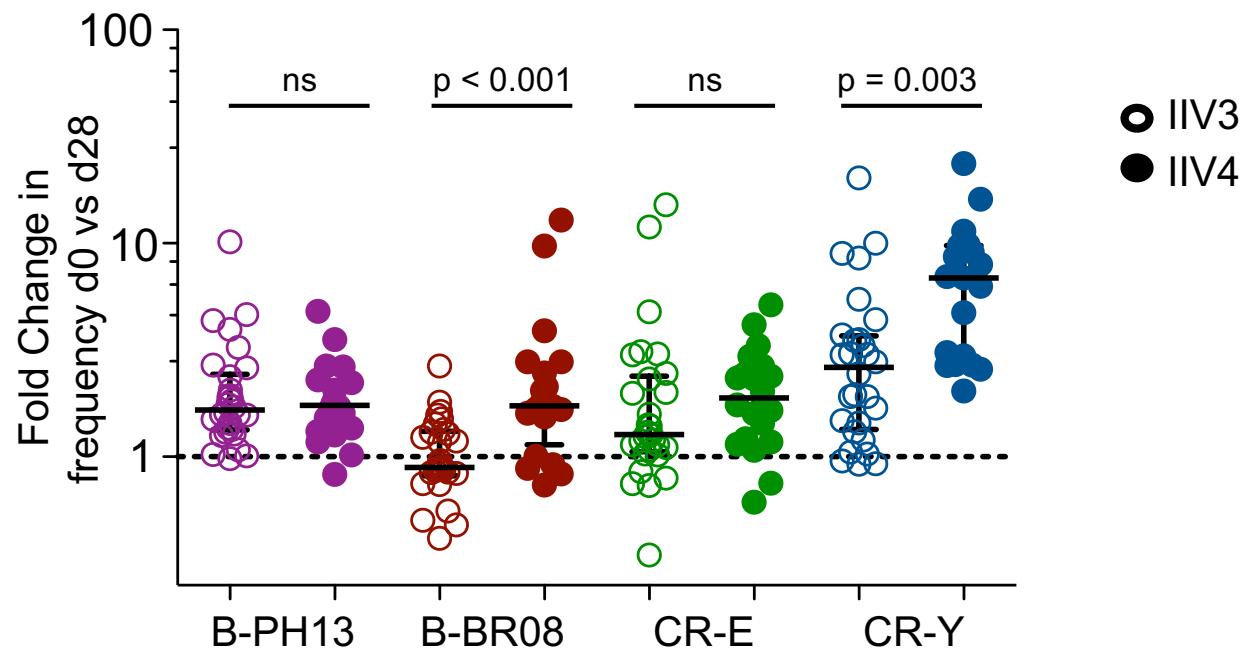
A (H1N1): an A/California/4/2009 (H1N1)pdm09 like virus
A (H3N2): an A/Hong Kong/4801/2014 (H3N2) like virus
B: a **B/Brisbane/60/2008** like virus
B: a **B/Phuket/3073/2013** like virus.



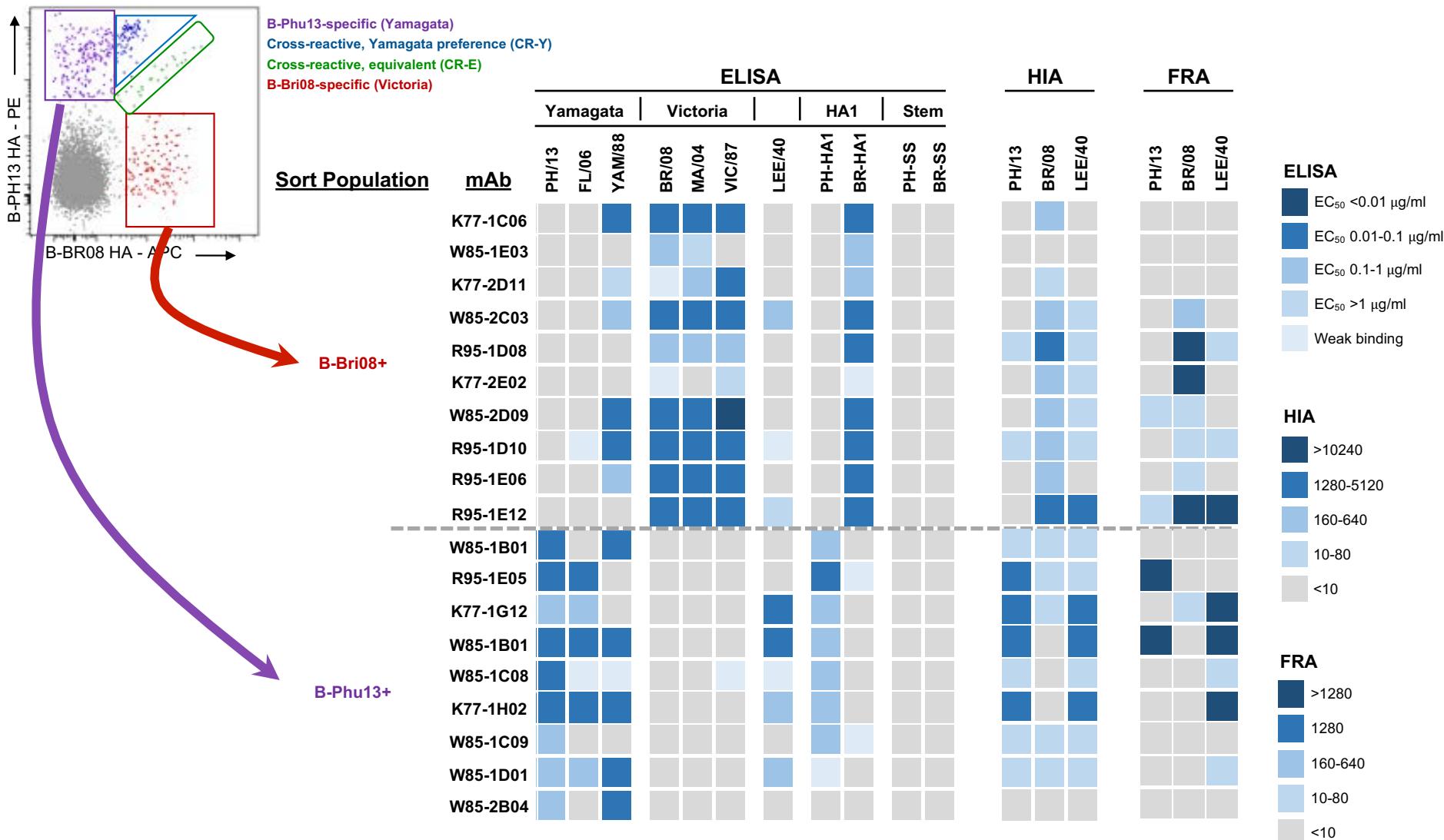
Seasonal vaccines drive the expansion of IBV cross-reactive B cells



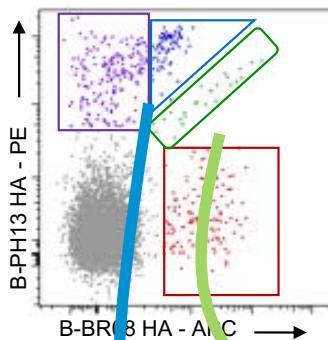
B-Phu13-specific (Yamagata)
Cross-reactive, Yamagata preference (CR-Y)
Cross-reactive, equivalent (CR-E)
B-Bri08-specific (Victoria)



“Strain-specific” B cells actually recognise pan-lineage HA



Highly cross-reactive B cells bind the IBV HA head and stem



B-Phu13-specific (Yamagata)
Cross-reactive, Yamagata preference (CR-Y)
Cross-reactive, equivalent (CR-E)
B-BR08-specific (Victoria)

Sort Population

mAb

W85-3F06

K77-1C02

R95-1H09

K77-1G03

R95-1B11

K77-1G12

R95-1C01

R95-1D05

K77-2B06

K77-2D11

R95-1E03

R95-1E05

K77-2H12

R95-1E07

R95-1F04

W85-1A07

R95-1H08

K77-1H05

W85-3E10

R95-1H09

K77-2C08

W85-3G03

K77-2D09

K77-2E07

R95-2A08

R95-2C02

R95-2G10

ELISA

Yamagata | Victoria | | HA1 | Stem

PH/13 FL/06 YAM/88 BR/08 MA/04 VIC/87 LEE/40 PH-HA1 BR-HA1 PH-SS BR-SS

HIA

PH/13 BR/08 LEE/40

FRA

PH/13 BR/08 LEE/40

ELISA

EC₅₀ <0.01 µg/ml

EC₅₀ 0.01-0.1 µg/ml

EC₅₀ 0.1-1 µg/ml

EC₅₀ >1 µg/ml

Weak binding

HIA

>10240

1280-5120

160-640

10-80

<10

FRA

>1280

1280

160-640

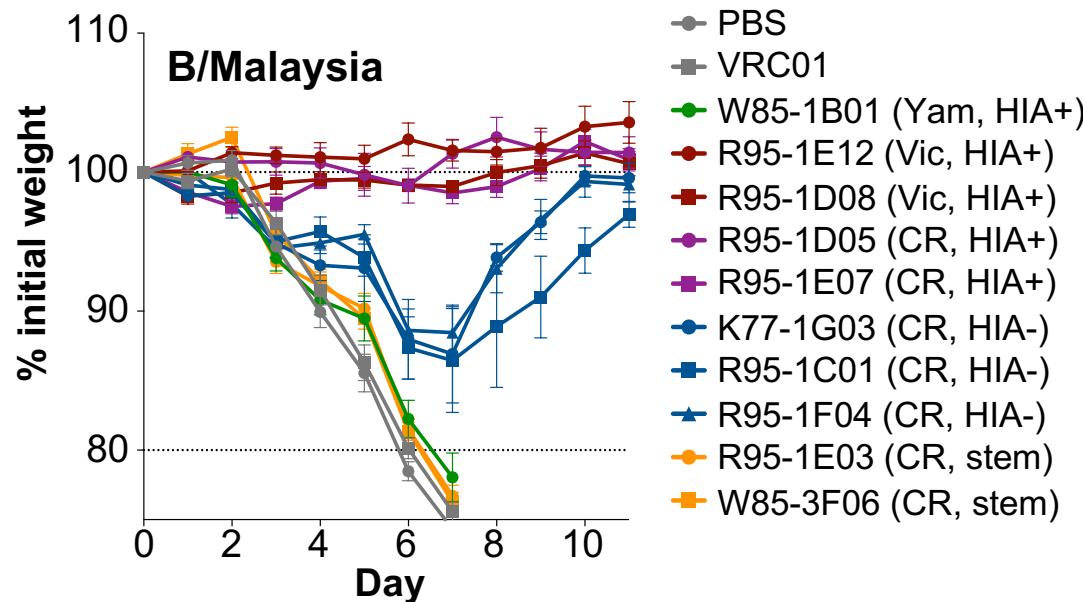
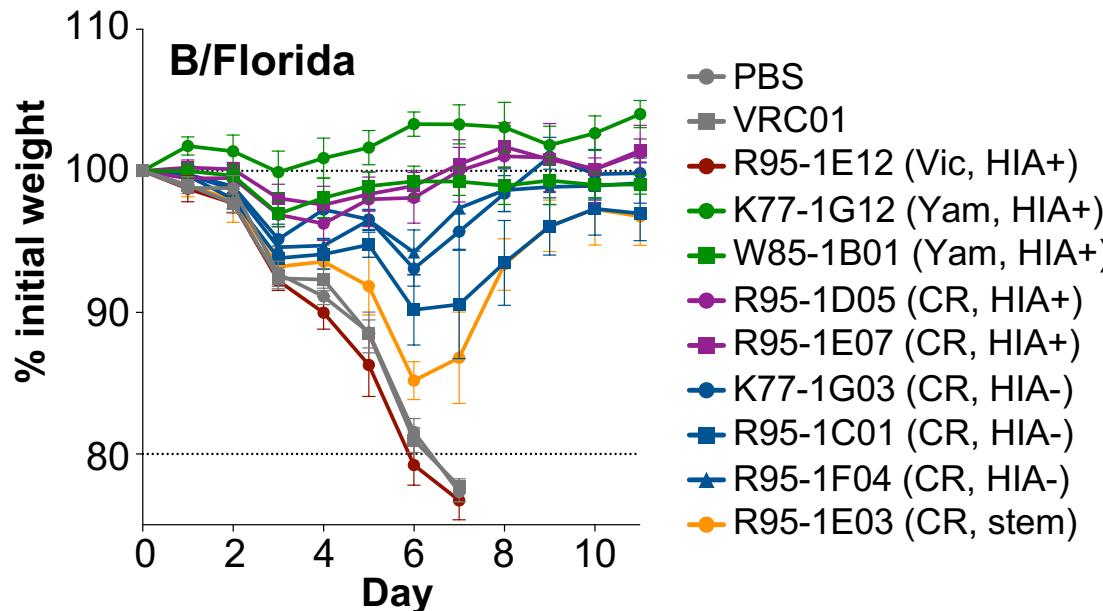
10-80

<10

Cross-reactive equivalent (CR-E)

Cross-reactive Yamagata (CR-Y)

Passive protection by human mAbs in IBV challenged mice

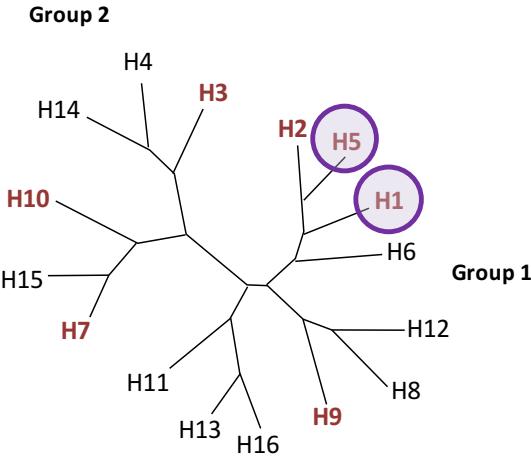


Summary

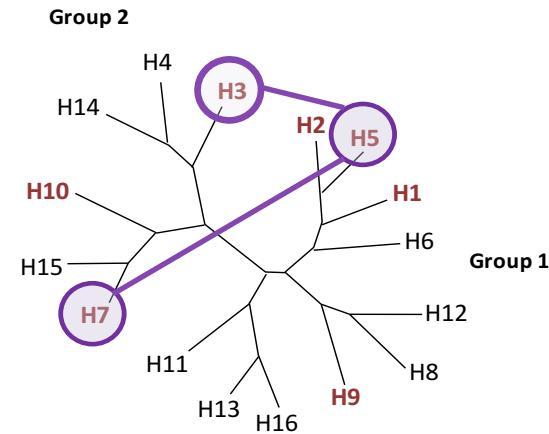
- IBV “strain-specific” memory B cells recognise pan-lineage HA (>25y antigenic drift)
 - Contrasts with IAV
- Cross-lineage B cells are common in seasonal vaccine recipients
 - CR-E B cells – often pan-IBV recognition
 - CR-Y B cells - more patchy
- Monoclonal antibodies derived from cross-lineage B cells
 - Protect mice in passive infusion studies
 - Can recognise HAI+ epitopes in the HA head
 - Often require Fc function for protection
 - Stem mAbs are poorly protective
- Both IIV3 and IIV4 drive expansion of IBV cross-reactive B cells
- **Cross-reactive B cell interrogation and isolation of monoclonal antibody lineages**
 - Potential to inform vaccine design
 - Identify novel immunotherapeutics

What about IAV?

inter-group1



group1/2



- Mostly stem
- Frequent
- Neutralising
- VH1-69 derived
- Age-associated increase in frequencies
- Inducible by immunisation (H5, pH1)

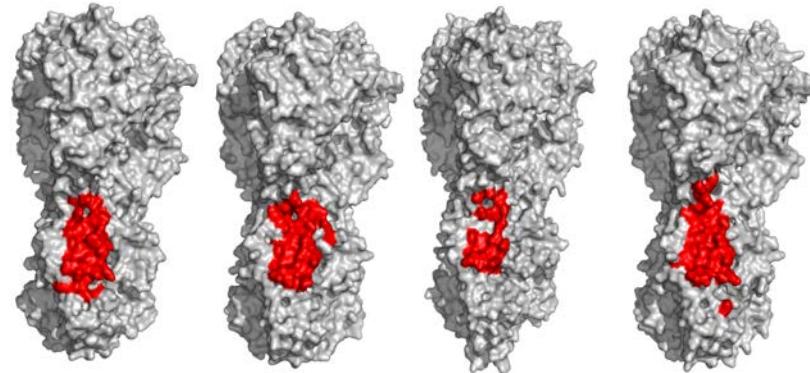
- Mostly stem
- Infrequent
- Neutralising
- Found in common "classes" between donors
- Inducible by immunisation (H5, H7)

Whittle et al. (2014) *J Virology*
Wheatley et al. (2015) *J Immunol*

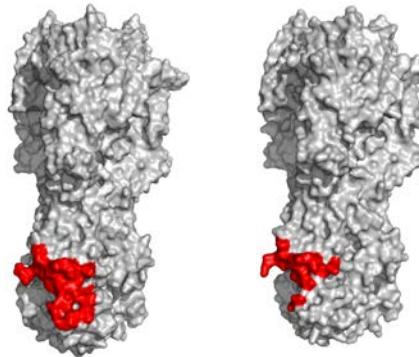
Joyce et al. (2016) *Cell*
Andrews et al. (2017) *Sci Immunol*

For IAV - the HA stem as conserved “supersite” of antibody recognition

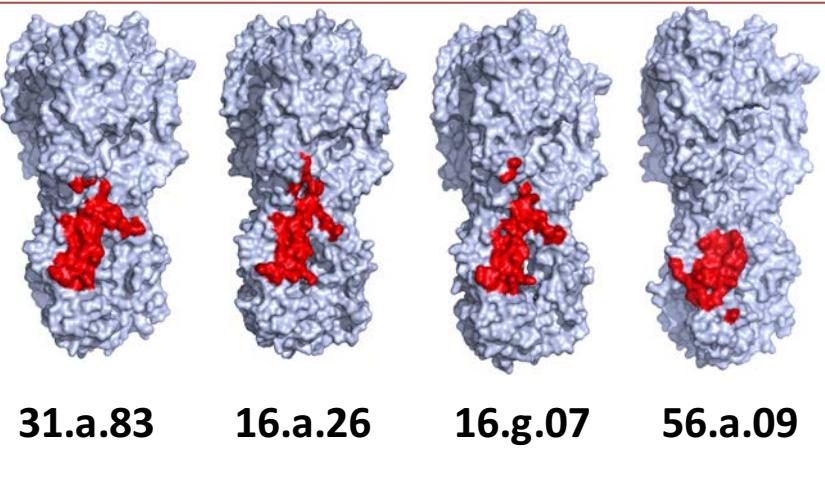
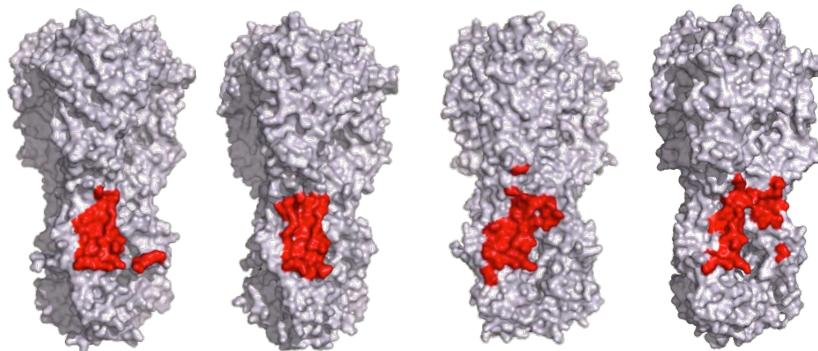
Pan-Group1 (mostly VH1-69)



Pan-Group2

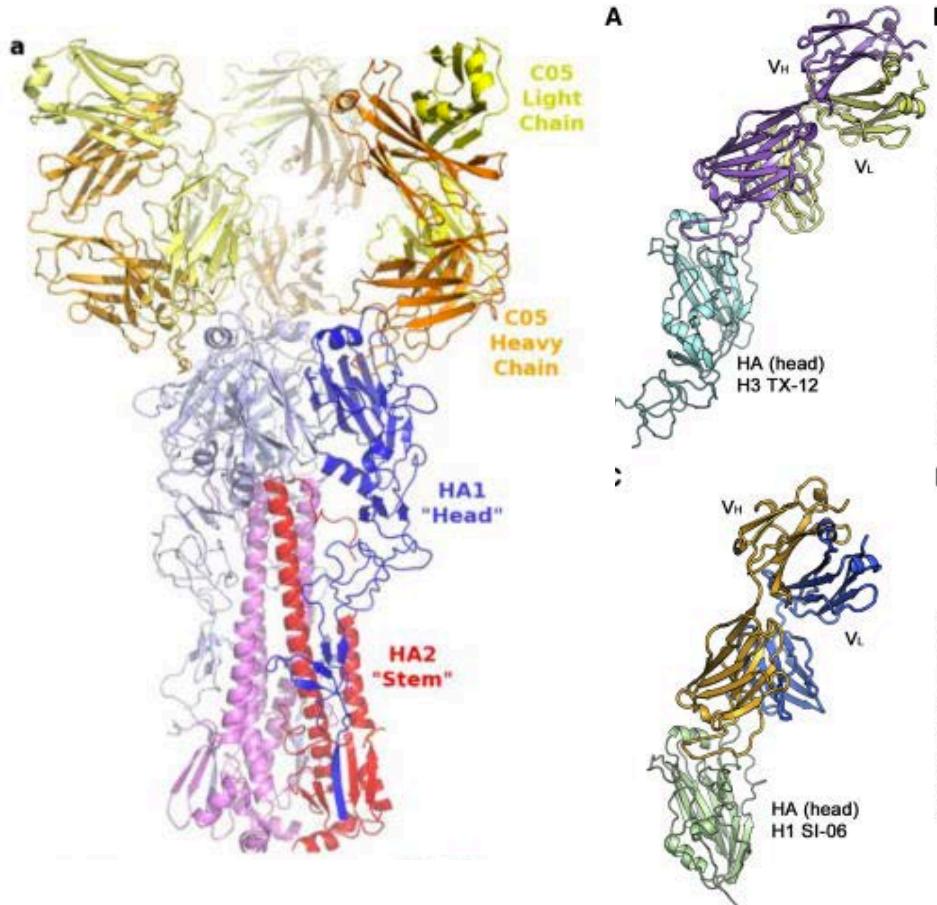


Inter-group

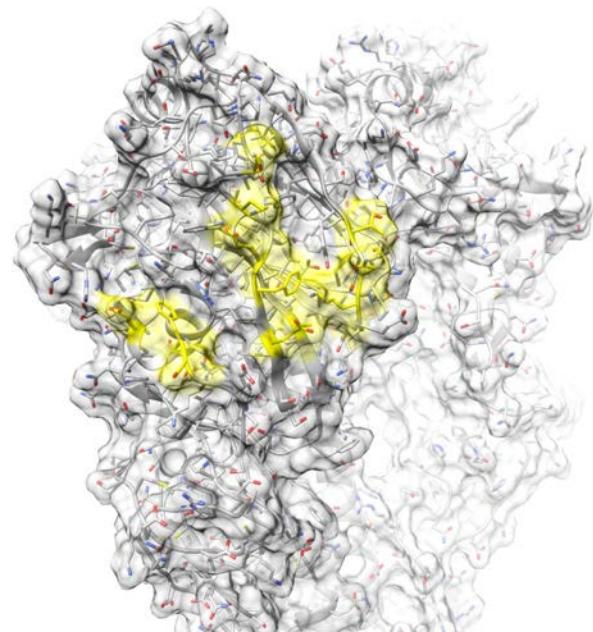


Cross-reactive, non-stem epitopes?

mAbs that mimic the SA receptor



Alternative sites in the HA head



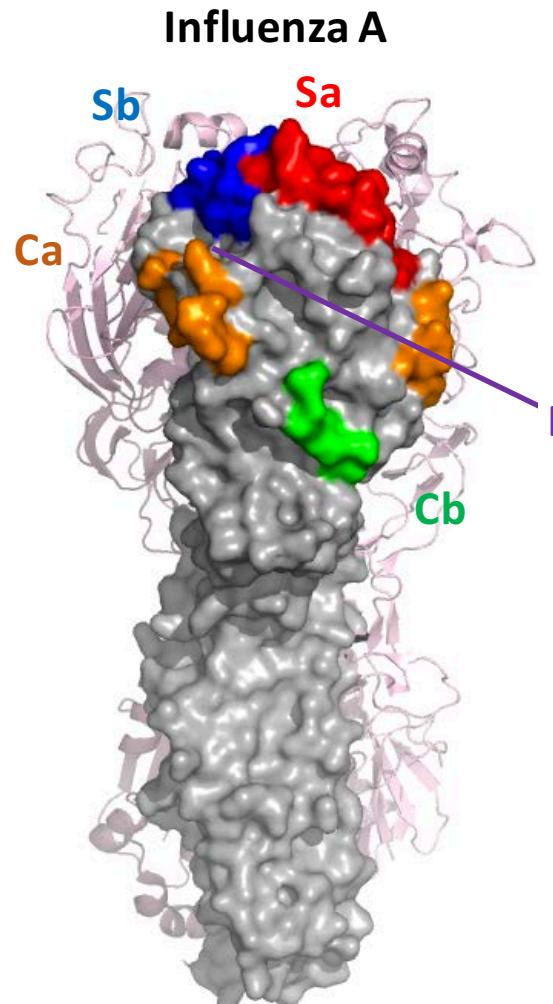
mAb 446D1
Pan-H1 activity

Ekiert et al. *Nature*. 2012

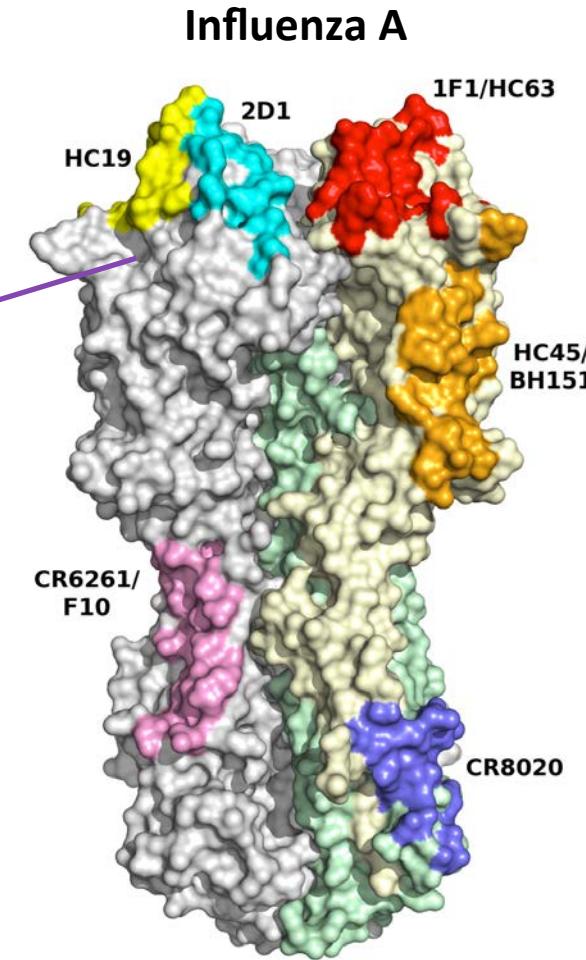
McCathey et al. *Immunity*. 2018

Kanekiyo et al. *Nat Immunol*. *In Press*

Moving on from classical epitopes to new targets (IAV)



Strain-specific



Cross-reactive

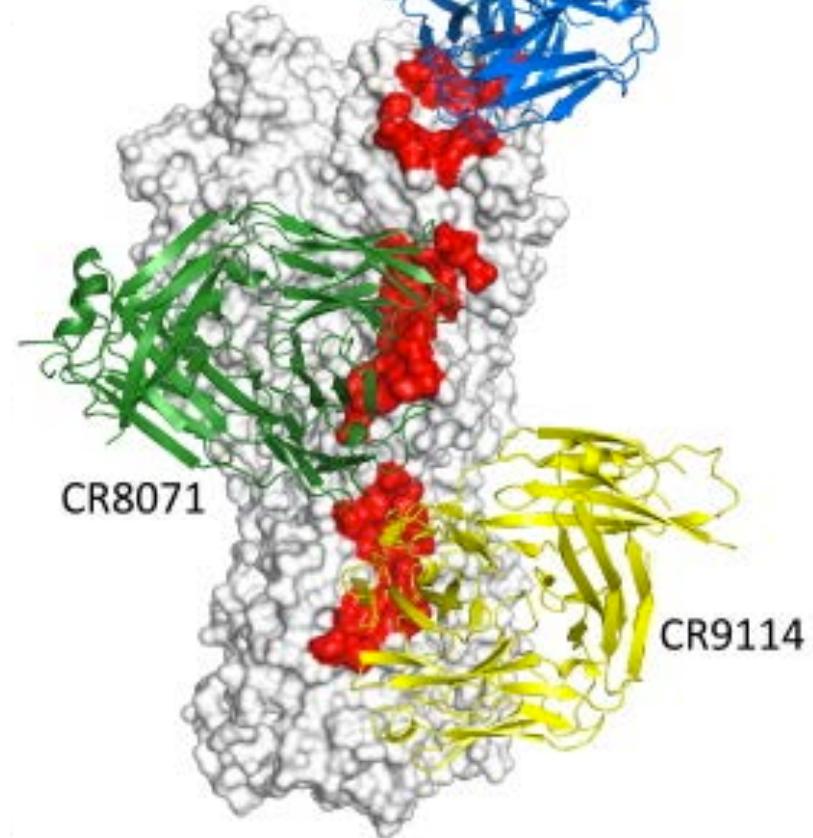
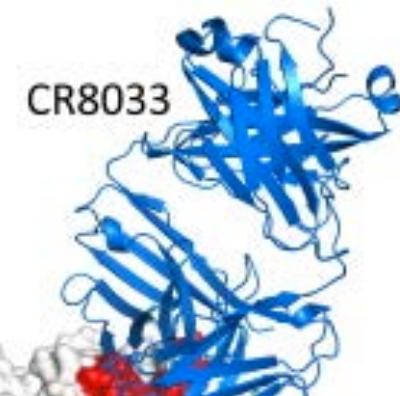
Moving on from classical epitopes to new targets (IBV)

Influenza B

HA1 (head)

HA2 (stem)

Strain-specific



Cross-reactive

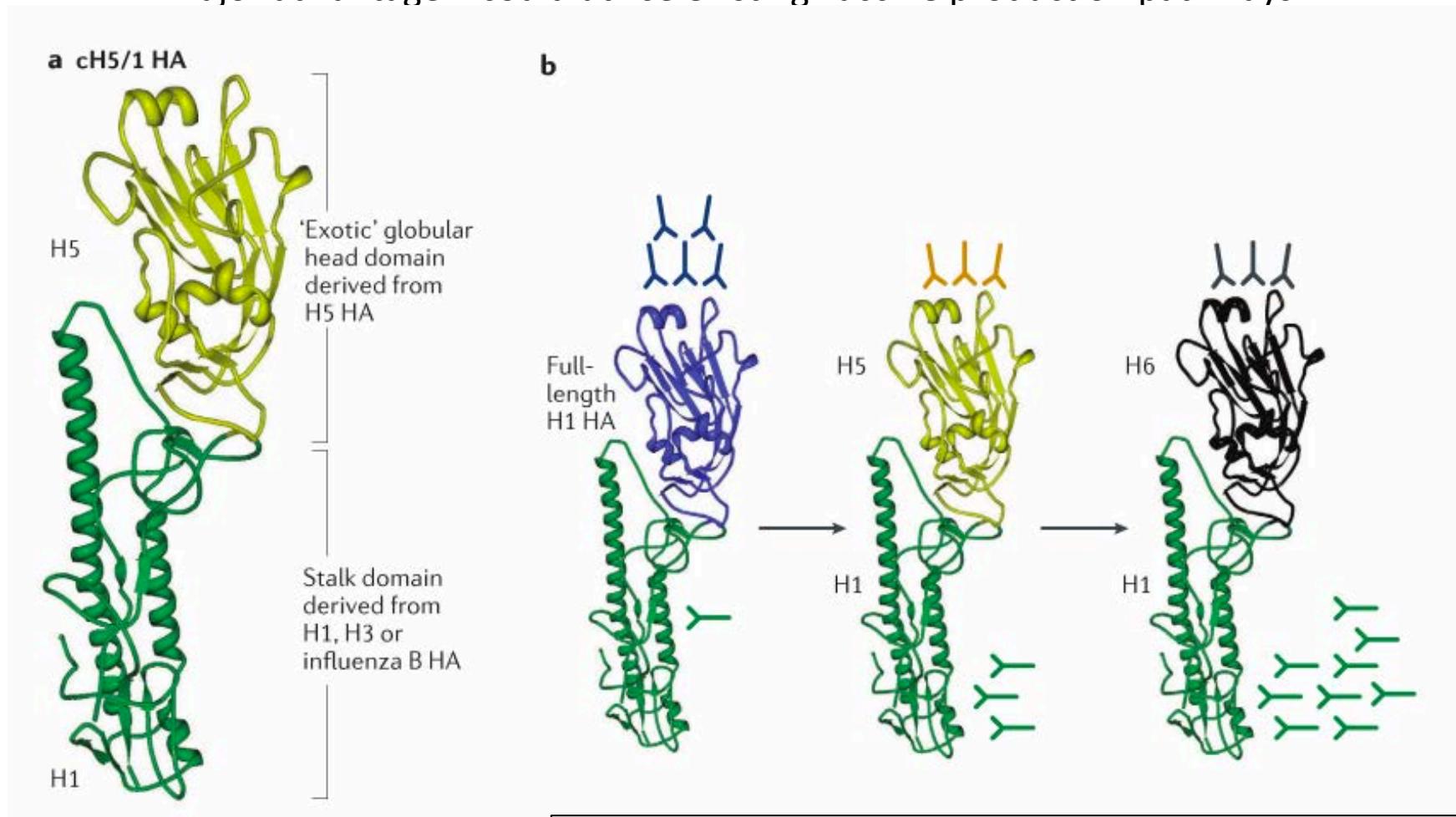
A strategy for HA-directed universal vaccines

1. Identify cross-reactive HA epitopes recognised by human antibody/B cells
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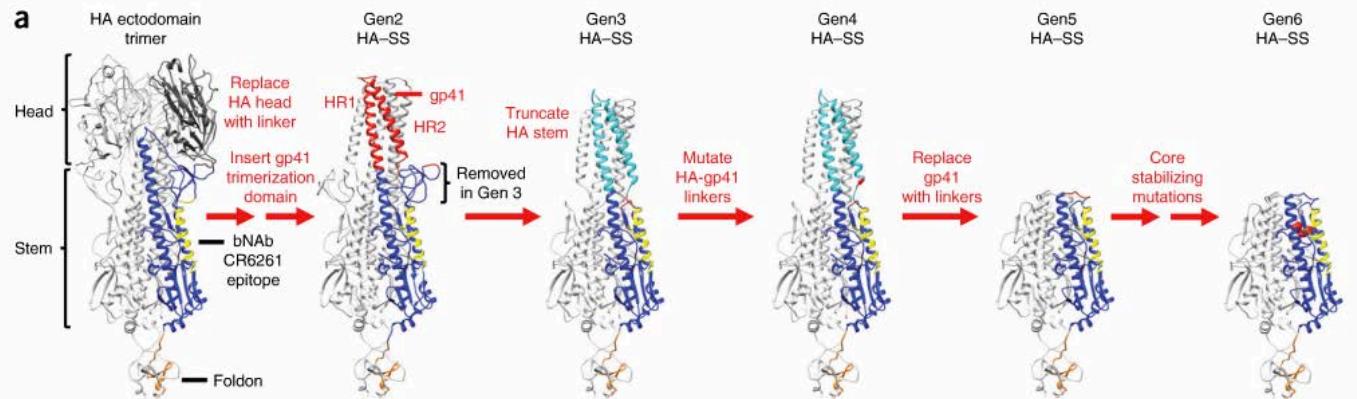
Vaccination with chimeric HA

- Strategies that maximise exposure of the HA stem by sequential vaccination with chimeric HA molecules bearing a common HA2 domain
- → Major advantage – could utilise existing vaccine production pathways



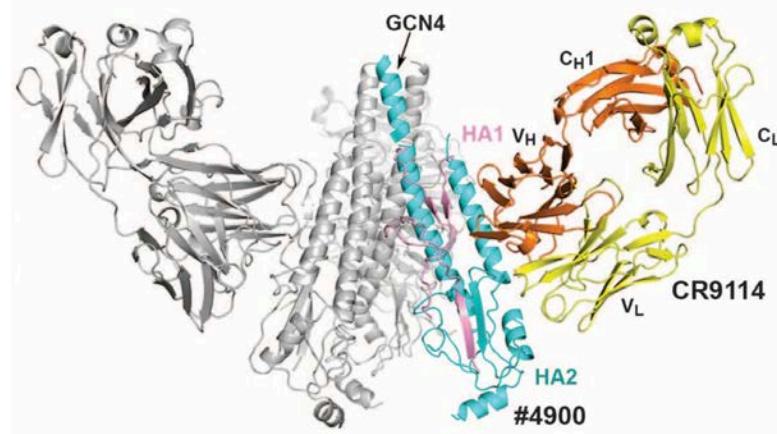
Hemagglutinin-stem nanoparticles generate heterosubtypic influenza protection

Hadi M Yassine^{1,6}, Jeffrey C Boyington^{1,6}, Patrick M McTamney^{1,5,6}, Chih-Jen Wei^{1,5,6}, Masaru Kanekiyo¹, Wing-Pui Kong¹, John R Gallagher², Lingshu Wang¹, Yi Zhang¹, M Gordon Joyce¹, Daniel Lingwood^{1,5}, Syed M Moin¹, Hanne Andersen³, Yoshinobu Okuno⁴, Srinivas S Rao^{1,5}, Audray K Harris², Peter D Kwong¹, John R Mascola¹, Gary J Nabel^{1,5} & Barney S Graham¹



A stable trimeric influenza hemagglutinin stem as a broadly protective immunogen

Antonietta Impagliazzo,^{1,*†} Fin Milder,^{1,§} Harmjan Kuipers,^{1,§} Michelle V. Wagner,^{2,||} Xueyong Zhu,^{3,‡} Ryan M. B. Hoffman,^{3,‡} Ruud van Meersbergen,^{1,§} Jeroen Huizingh,^{1,§} Patrick Wanningen,^{1,§} Johan Verspuij,^{1,§} Martijn de Man,^{1,§} Zhaqing Ding,^{2,||} Adrian Apetri,^{1,†} Başak Kükrer,^{1,†} Eveline Sneekes-Vriese,¹ Danuta Tomkiewicz,^{1,†} Nick S. Laursen,^{3,¶} Peter S. Lee,³ Anna Zakrzewska,^{1,§} Liesbeth Dekking,^{1,§} Jeroen Tolboom,^{1,§} Lisanne Tettero,^{1,§} Sander van Meerten,^{1,§} Wenli Yu,³ Wouter Koudstaal,^{1,†} Jaap Goudsmit,^{1,†} Andrew B. Ward,³ Wim Meijberg,^{1,§} Ian A. Wilson,^{3,**} Katarina Radošević^{1,§}

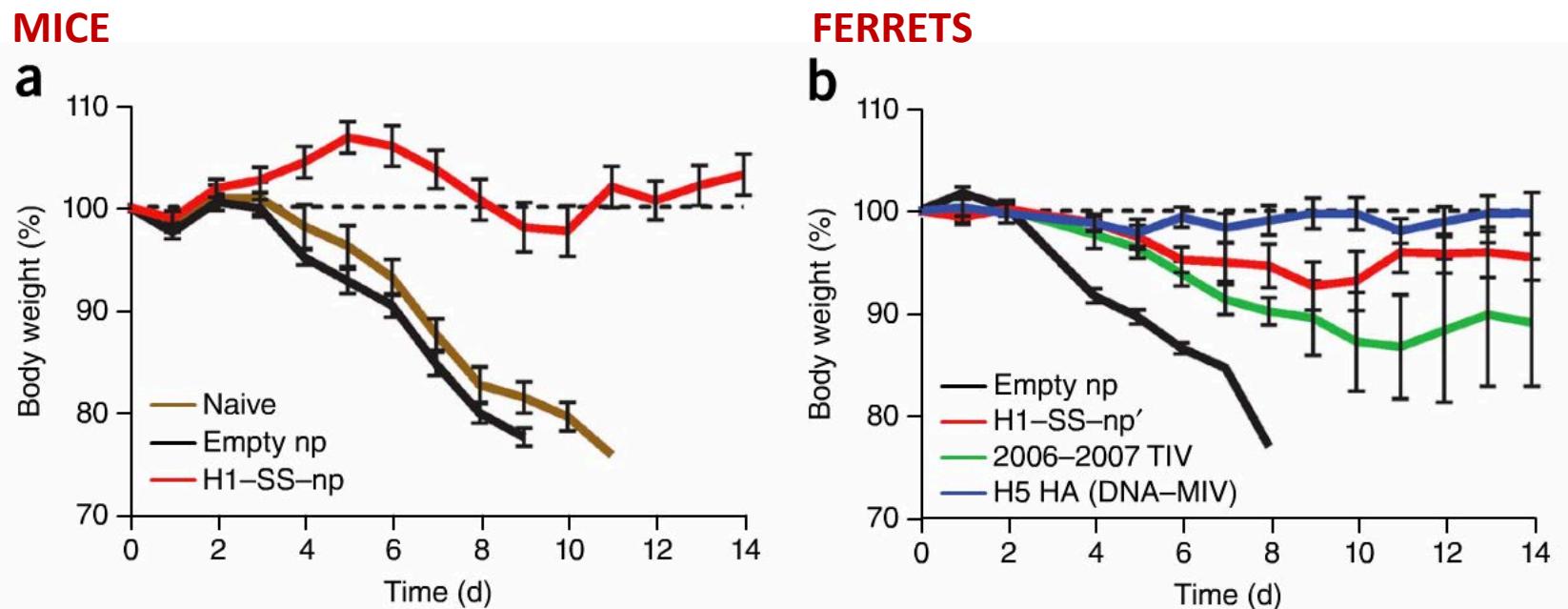


Yassine et al. (2015) *Nat Med* 21(9):1065-70

Impagliazzo et al. (2015) *Science* 349(6254):1301-6

Hemagglutinin-stem nanoparticles generate heterosubtypic influenza protection

Hadi M Yassine^{1,6}, Jeffrey C Boyington^{1,6}, Patrick M Tamney^{1,5,6}, Chih-Jen Wei^{1,5,6}, Masaru Kanekiyo¹, Wing-Pui Kong¹, John R Gallagher², Lingshu Wang¹, Yi Zhang¹, M Gordon Joyce¹, Daniel Lingwood^{1,5}, Syed M Moin¹, Hanne Andersen³, Yoshinobu Okuno⁴, Srinivas S Rao^{1,5}, Audray K Harris², Peter D Kwong¹, John R Mascola¹, Gary J Nabel^{1,5} & Barney S Graham¹



- Vaccination with “stabilised stem” nanoparticles derived from seasonal H1N1 (A/New Caledonia/20/99) protected against lethal H5N1 (VN) challenge in mice and ferrets

Summary – Human cross-reactive HA epitopes

- Humans make subdominant but highly cross-reactive humoral responses to influenza
- Cross-reactive memory B cells recognise epitopes in both the head (HA1) and stem (HA2) domain
- Can mediate a range of effector functions – ADCC etc.
- To date, the broadest neutralising antibodies all bind in the HA stem
- Epitopes and B cell lineages remarkably conserved between antibodies/subjects
 - Convergent structures
 - Convergent genetic elements
- Breadth can extend to:
 - IAV - diverse Group1 and Group2 viruses
 - IBV – both antigenic lineages

Summary – HA-based UIV approaches

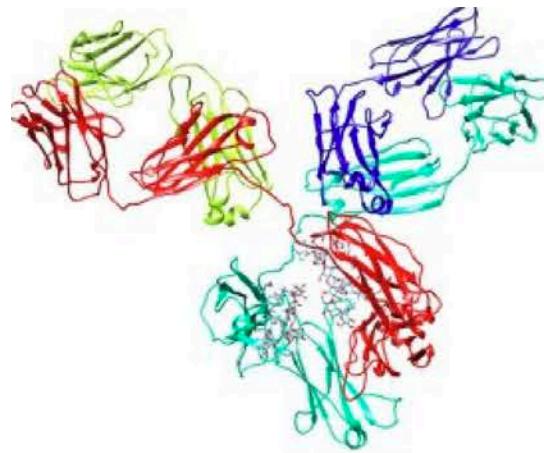
- Both chimeric HA and headless stem only constructs
 - Commencing clinical testing
- **Question - Translatability of animal immunisation models to humans**
 - Clarify the influence of pre-existing baseline immunity in humans
 - Development of animal models to better account for influenza immunity
- **Question - Can dominant HAI+ antibody responses be avoided/subverted?**
- **Question – Is HA stem antibody at all protective?**
 - Acquisition
 - Severity

The case for monoclonal antibody therapeutics/prophylactics

There is a need for novel agents for:

- clinical treatment of severe influenza infection
- pandemic prophylaxis

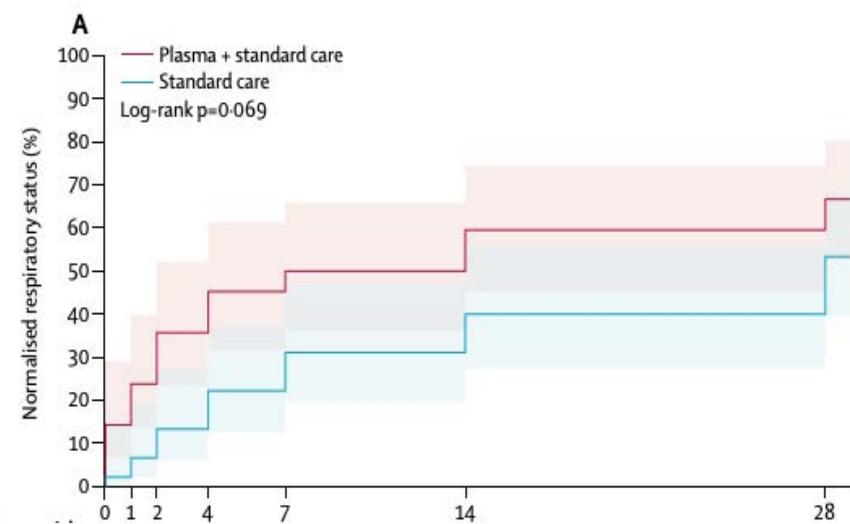
What about mAbs?



- **Advantages**
 - Rapid activity
 - High tolerability
 - Low/no resistance
 - Long biological half-life (advantage for prophylaxis)
- **Disadvantages**
 - Cost (getting lower but still high)
 - Need to account for viral diversity

Anti-influenza mAbs – are they actually protective?

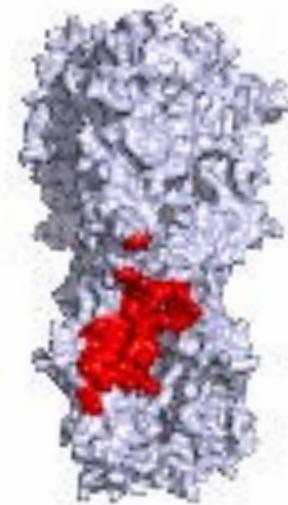
- 1918 influenza pandemic – treatment with plasma, serum or blood
 - Reduced case-fatality rate – (16% vs 37%) – Luke et al. 2006 *Ann Intern Med*
- Immune plasma for the treatment of severe influenza
 - Some potential clinical benefit
 - Beigal et al. 2017 *Lancet Respir Med*
- INSIGHT FLU005
 - Assessing flu-IVIG for clinical benefit
- Human monoclonal antibodies – extensively characterised in mice and ferrets
 - Effective treatment – Up to 3 days post-infection
 - Prophylactic protection against seasonal and high path avian viruses
 - Generally utilise Fc-FcR interactions (ADCC, ADP) for protection



Anti-influenza mAbs – human clinical trials

MEDI8852 (Medimmune) – Group1/Group2 IAV stem

- reduced symptoms during natural infection
- Ali et al. 2018 AAC



MHAA4549A

VIS410 (Visterra) - Group1/Group2 IAV stem

- Reduction in viral shedding during pH1N1 challenge : Sloan et al. Options IX

CR9114/CR6261/CR8020 (Crucell) - Group1 and/or Group2 IAV stem

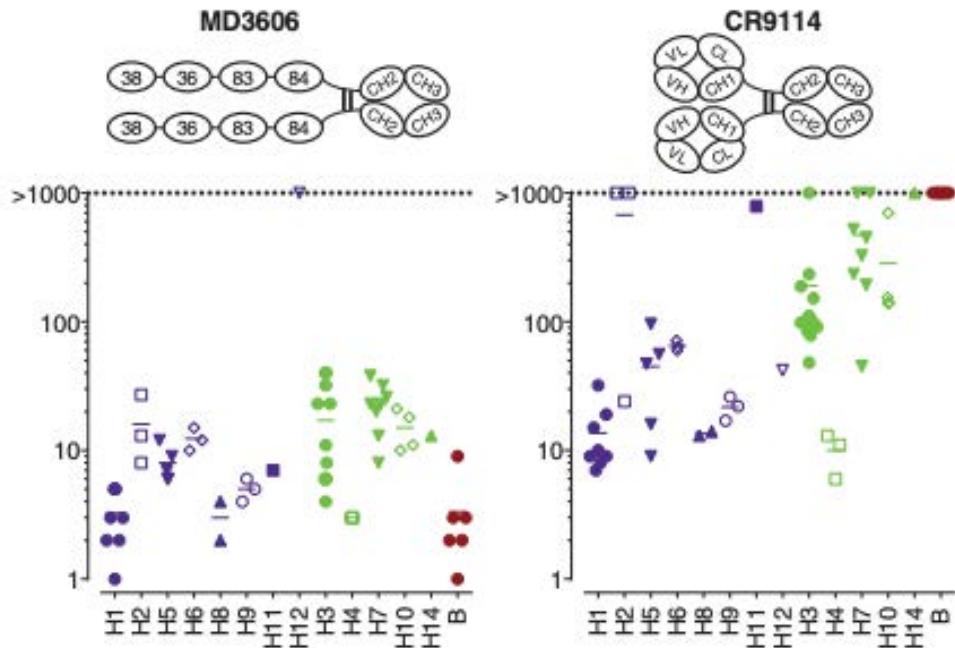
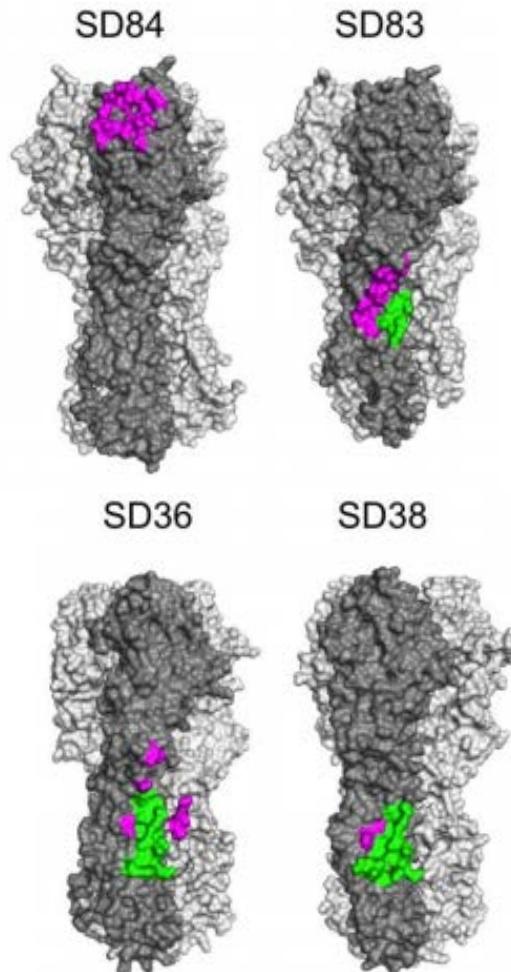
- Trials pulled or not reported

TCN-032 (Theraclone) - anti-M2e

- lower symptom scores and shedding during H3N2 challenge : Ramos et al. 2015

Universal protection against influenza infection by a multidomain antibody to influenza hemagglutinin

Nick S. Laursen^{1*}, Robert H. E. Friesen^{2†}, Xueyong Zhu¹, Mandy Jongeneelen³,
Sven Blokland³, Jan Vermond⁴, Alida van Eijgen⁴, Chan Tang³, Harry van Diepen⁴,
Galina Obmolova², Marijn van der Neut Kolschoten³, David Zuidgeest³,
Roel Straetemans⁵, Ryan M. B. Hoffman¹, Travis Nieuwsma¹, Jesper Pallesen¹,
Hannah L. Turner¹, Steffen M. Bernard¹, Andrew B. Ward¹, Jinquan Luo²,
Leo L. M. Poon⁶, Anna P. Tretiakova^{7‡}, James M. Wilson⁷, Maria P. Limberis⁷,
Ronald Vogels³, Boerries Brandenburg³, Joost A. Kolkman^{8§}, Ian A. Wilson^{1,9§}



Summary – anti-influenza mAbs

- Animal trials consistently show human mAbs have promise
- Evidence of clinical benefit for mAb treatment in humans still sparse
 - Natural infection - some symptom alleviation
 - Experimental challenge models - some reductions in viral burden
 - Hampered by poor reporting
 - Larger trials are ongoing
- No demonstration of prophylactic effect to date in humans
- New mAbs or antibody-engineering technologies
 - Opportunity for using potent neutralising antibodies while maintaining effective breadth
 - Opportunity to increase biological half-lives, respiratory tract delivery

Acknowledgments



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Oanh Nguyen
Marios Koutsakos
Kanta Subbarao
Aeron Hurt
Danni, Malet



National Institute of
Allergy and
Infectious Diseases

*Dale and Betty Bumpers
Vaccine Research Center*



M Gordon Joyce

Barney Graham
Masaru Kanekiyo



Hadi Yassine

Clinical trial teams and participants

Funding: CONVERGENT
BIO-NANO SCIENCE
& TECHNOLOGY



Australian Government

National Health and Medical Research Council